# Extraction of 25-hydroxy Vitamin D from Serum Using ISOLUTE® PLD+ prior to LC-MS/MS Analysis

This application note describes the extraction of 25-hydroxy vitamin D from serum, prior to LC-MS/MS analysis.

Figure 1. Structures of 25-hydroxy Vitamin D

#### Introduction

ISOLUTE® PLD+ Protein and Phospholipid Removal plates offer a substantial improvement in extract cleanliness compared to traditional protein precipitation techniques for bioanalytical sample preparation.

This application note describes a simple, effective ISOLUTE° PLD+ protocol for the extraction of 25-hydroxy vitamin D from serum, demonstrating high, reproducible analyte recoveries with low protein and phospholipid content in the extracts.

# **Analytes**

25-hydroxy vitamin D2, 25-hydroxy vitamin D3 and d6-25-hydroxy vitamin D3 as the internal standard.

# Sample Preparation Procedure

Format: ISOLUTE® PLD+ Protein and Phospholipid Removal plate, part number 918-0050-P01

## Sample Pre-treatment

Add 10  $\mu$ L of ISTD (equivalent to 30 ng/mL) to the serum sample. Mix. Allow to stand for ~1 hour for binding to occur.

## **Solvent Application**

Apply 400  $\mu$ L of Acetonitrile (MeCN) to each well of the ISOLUTE PLD+ plate.

## **Sample Application**

Add 100  $\mu$ L of serum with ISTD and mix thoroughly via repeat aspirate/dispense steps.

## **Analyte Elution**

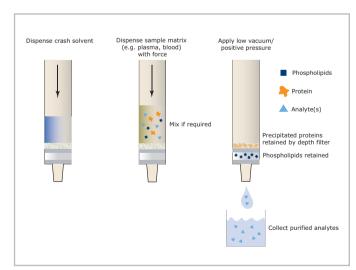
Apply vacuum -o.2 bar or 3 psi positive pressure for approximately 5 minutes. For highly particulate laden samples increased pressure or vacuum conditions may be required.

#### **Post Extraction**

Dry the extract in a stream of air or nitrogen using a SPE Dry (40 °C at 40 L/min) or TurboVap (40 °C at 1.0 bar).

#### Reconstitution

Apply 100  $\mu L$  of 30/70 2 mM Ammonium Formate, 0.1% formic acid aq/MeOH.



 $\textbf{Figure 2.} \ \, \textbf{Typical ISOLUTE}^{\circ} \ \, \textbf{PLD+procedure}$ 



## **UPLC** Conditions

#### Instrument

Waters Acquity UPLC (Waters Assoc., Milford, MA, USA)

#### Column

ACE EXCEL 2 C18-PFP, 100 mm x 2.1 mm id 2  $\mu$ m, (ACT, UK)

#### **Mobile Phase**

A: 2 mM ammonium formate/o.1% formic acid (aq)
B: 2 mM ammonium formate/o.1% formic acid/MeOH

#### **Flow Rate**

o.4 mL/min

Table 1. UPLC Gradient Conditions.

| Time | %A | %В  | Curve |
|------|----|-----|-------|
| 0    | 25 | 75  | 1     |
| 3    | 0  | 100 | 6     |
| 4    | 25 | 75  | 11    |

**Curve 11:** Conditions in line initiated immediately once time passed. i.e. 25:75 resumed at 4 minutes.

Curve 6: Linear Gradient

## **Injection Volume**

15 μL (partial loop with overfill)

#### Sample Temperature

20 °C

#### **Column Temperature**

40 °C

**Note:** alternative UPLC conditions may be suitable. Check for good chromatographic separation of isobaric interferences to ensure accurate analyte quantitation

# Mass Spectrometry Conditions

#### Instrument

Quattro Premier XE triple quadrupole mass spectrometer (Waters Assoc., Manchester, UK) equipped with an electrospray interface for mass analysis

#### **Desolvation Temperature**

450 °C

#### **Ion Source Temperature**

150 °C

#### **Collision Cell Pressure**

3.76 e<sup>-3</sup> mbar

Positive ions acquired in the multiple reaction monitoring (MRM) mode:

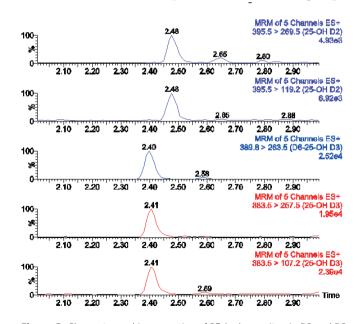
**Table 2.** Quattro Premier XE MRM parameters (Qualifier ion details shown in parenthesis).

| Analyte                 | MRM Transition                   | Cone V | Collision<br>Energy eV |
|-------------------------|----------------------------------|--------|------------------------|
| 25-OH D <sub>2</sub>    | 395.5 > 269.5<br>(395.5 > 119.2) | 30     | 18<br>26               |
| 25-OH D <sub>3</sub>    | 383.5 > 257.5<br>(383.5 > 107.2) | 30     | 17<br>25               |
| d6-25-OH D <sub>3</sub> | 389.6 > 263.5                    | 30     | 16                     |

# Results

#### Chromatography

Good chromatographic separation of 25-hydroxy vitamin D2 and D3 was achieved in less than 3 minutes, as shown in **Figure 3.** 



**Figure 3.** Chromatographic separation of 25-hydroxy vitamin D2 and D3 from Chromsystems calibrated serum at 14.8 and 19.6  $\,$  ng/mL respectively. ISTD at 30  $\,$  ng/mL.



# Recovery

Serum and stripped serum was spiked at various concentrations from 2–100 ng/mL. High reproducible recoveries > 70% with corresponding RSDs < 10% were demonstrated. Typical recovery data is shown in **Figure 4.** 

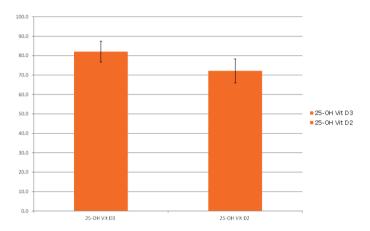


Figure 4. Recovery profile for 25-hydroxy vitamin D extracted at 50 ng/mL.

# PBS/BSA Calibration Curves

Calibration curves were generated using PBS/BSA spiked at concentrations from 1–100 ng/mL. Good coefficients of determination were obtained for 25-hydroxy vitamin D2 and D3 ( $r^2 > 0.99$ ).

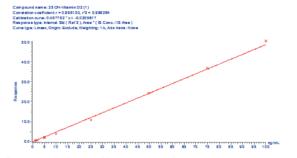
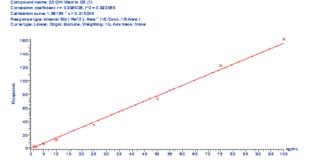


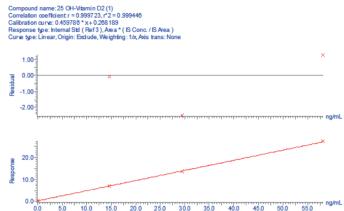
Figure 5. PBS/BSA Calibration Curve for 25-OH vitamin D2 constructed from 1–100  $\mbox{ng/mL}.$ 



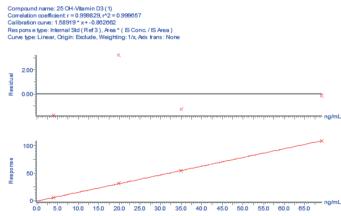
**Figure 6.** PBS/BSA Calibration Curve for 25-OH vitamin D3 constructed from 1–100 ng/mL.

# **Chromsystems Calibration Curves**

Curves were also generated using calibrated serum standards (obtained from Chromsystems) spiked at concentrations from o-69 ng/mL. Good coefficients of determination were obtained for 25-hydroxy vitamin D2 and D3 ( $r^2 > 0.99$ ).



**Figure 7.** Chromsystems Calibrated Serum Curve for 25-OH vitamin D2 constructed from 4-69 ng/mL.



**Figure 8.** Chromsystems Calibrated Serum Curve for 25-OH vitamin D3 constructed from 0-58 ng/mL.



# DEQAS External Quality Assessment Scheme

Final method testing was performed for 5 DEQAS serum samples extracted alongside the Chromsystems calibrators using the optimized method. The DEQAS criteria for acceptable performance is that at least 80% of results should fall within + or - 25% of the All Laboratory Trimmed Mean. Method performance is shown in Table 3. Units are quoted as ng/mL. All values fall within the accepted criteria.

Table 3. DEQAS 25-OH vitamin D results obtained using optimum method.

| DEQAS<br>Sample I.D. | DEQAS<br>LC/MS Mean | ISOLUTE°<br>PLD+ |
|----------------------|---------------------|------------------|
| 451                  | 12.9                | 14.5             |
| 452                  | 46.7                | 49.1             |
| 453                  | 26.6                | 28.9             |
| 454                  | 21.4                | 25.3             |
| 455                  | 22.2                | 23.7             |

# **Ordering Information**

| Part Number    | Description  | Quantity |
|----------------|--|----------|
| 918-0050-P01   | ISOLUTE® PLD+ Fixed Well Plate                           | 1        |
| 121-9600       | Biotage® VacMater™-96<br>Sample Processing Manifold      | 1        |
| PPM-96         | Biotage® PRESSURE+ 96<br>Positive Pressure Manifold      | 1        |
| SD-9600-DHS-EU | Biotage® SPE Dry Sample<br>Concentrator System 220/240 V | 1        |
| SD-9600-DHS-NA | Biotage® SPE Dry Sample<br>Concentrator System 100/120 V | 1        |
| C103263        | TurboVap*96,<br>Evaporator 100/120V                      | 1        |
| C103264        | TurboVap® 96,<br>Evaporator 220/240V                     | 1        |

# **Additional Notes**

## **Buffer Preparation**

- 1. 2 mM ammonium formate/0.1% formic acid (aq): Weigh 0.12612 g and dissolve in H<sub>2</sub>O. Add 1 mL of formic acid and make up to 1 L in H<sub>2</sub>O.
- 2. 2 mM ammonium formate/0.1% formic acid/MeOH: Weigh 0.12612 g and dissolve in MeOH. Add 1 mL of formic acid and make up to 1 L in MeOH.

#### **Processing Conditions**

Positive Pressure: Process at approximately 3 psi.

Vacuum Processing: Process at approximately -0.2 bar.

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