

Peptide Mapping of Adeno-Associated Virus Capsid Proteins

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Overview

The use of Adeno-Associated Viruses (AAV) as an effective vector for the delivery of ssDNA in gene therapy, is well established and continues to grow.

As with any protein, viral proteins (VPs) are subject to post-translational modifications that can impact stability, capsid assembly, and overall efficacy. As a result, peptide mapping can be used to evaluate the presence and identification of these modifications.

At first glance AAVs present additional complexity not encountered with typical biologics. The viral capsid is a sixty-protein icosahedron comprised of three capsid, or viral proteins, non-covalently assembled in a ratio of 1:1:10 (VP1:VP2:VP3). However, a traditional peptide digest approach is sufficient to yield expected results. Although given the sequence homology, variation of signal intensity for peptides with overlapping and non-overlapping sequences should be anticipated.

In this application note, we profile capsid protein peptides from AAV5 and AAV9, digested with trypsin using a Biozen™ Peptide XB-C18 reversed phase column. We show the Biozen Peptide XB-C18 column provides excellent peak capacity and separation with a high level of sequence coverage.

Sample Preparation

Step	Description		
Sample:	$20~\mu\text{L}$ AAV with concentration 2E13 vg/mL		
Denaturation:	50 % v/v Trifluoroethanol		
Reduction:	2 mM Final Tris (2-carboxyethyl) Phosphine		
Incubation:	70 °C for 30 minutes		
Alkylation:	2 mM Final Iodoacetimide		
Incubation:	22 °C for 30 minutes, in the dark		
Dilution:	4.5X with 100 mM Ammonium Bicarbonate, pH 8.25		
Digestion:	1:25 w/w Trypsin		
Incubation:	37 °C for 18 hours		
Quench:	0.4 % v/v with 10 % Formic Acid		
Concentration:	SpeedVac to approx. 25 % of final reaction volume		
Resuspension:	Add equal volume of 25 mM Ammonium Bicarbonate, pH 8.25		

LC Conditions

Column: Biozen 2.6 µm Peptide XB-C18

Dimension: 150 x 2.1 mm **Part No.:** <u>00F-4768-AN</u>

Mobile Phase: A: 0.1 % Formic Acid in Water

B: 0.1 % Formic Acid in Acetonitrile

Gradient:	Time (min)	%E
	0	2
	50.5	60
	55.5	60
	60	90
	63	90
	63.1	2
	70	2

Flow Rate: 0.3 mL/min

Injection Volume: 1 μL Temperature: 60°C

Instrument: Vanquish™ UHPLC

Detection: MS

Detector: Q Exactive[™] Orbitrap[™] Plus

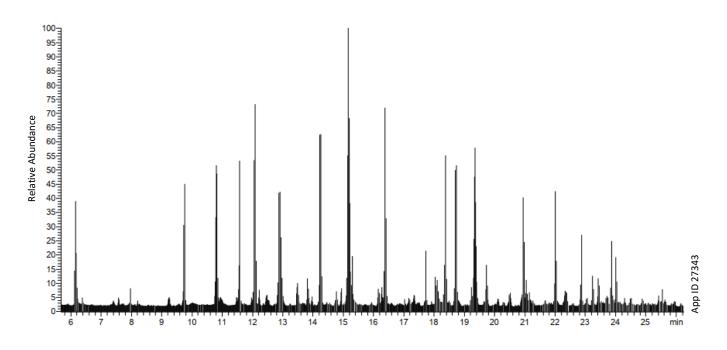
MS Conditions

Scan Type: Full MS
Resolution: 70,000
AGC Target: 3e6 ms
Maximum IT: 200 ms

Scan Range: 200 to 2000 m/z

Scan Type: MS2 Resolution: 70,000 AGC Target: 1e5 ms Maximum IT: 200 ms

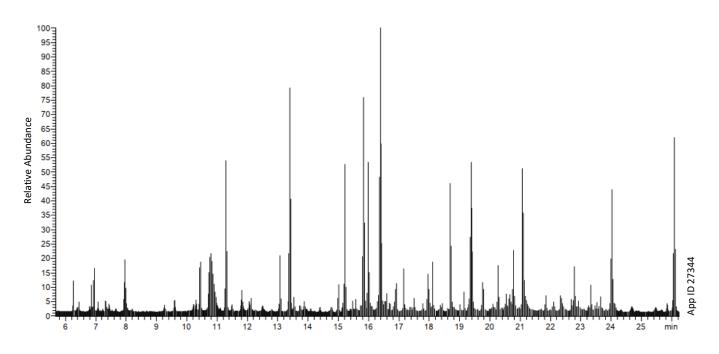
Figure 1. Peptide Mapping of AAV5 VPs, at a Concentration of 6.7E12 vg/mL (Approx. 55 ng/μL).



Protein Name	# Peptides	% Coverage	# AAs
>tr Q9YIJ1 Q9YIJ1_9VIRU Capsid protein OS=adeno-associated virus 5 OX=82300 GN=cap PE=1 SV=1	193	93.6	723

>tr/Q9YIJ1/Q9YIJ1_9YIRU Capsid proteim OS-ademo-associated virus 5 OX-82300 GN-cap PE-1 SV-1

Figure 2. Peptide Mapping of AAV9 VPs, at a Concentration of 6.7E12 vg/mL (Approx. 55 ng/μL).



Protein Name	# Peptides	% Coverage	# AAs
>tr Q6JC40 Q6JC40_9VIRU Capsid protein VP1 OS=Adeno-	205	92 7	768
associated virus 9 OX=235455 GN=cap PF=1 SV=1	203	32.7	700

>tr/Q6JC40/Q6JC40_9VIRU Capsid proteim VP1 OS=&demo-associated virus 9 OX=235455 GN=cap PE=1 SV=1

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