

# Enhancing the Scope for Screening Protein-Ligand Interactions by Non-Covalent Mass Spectrometry

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Inspired by **patients.**  
Driven by **science.**

# Non-Covalent Mass Spectrometry in NCE Drug Discovery

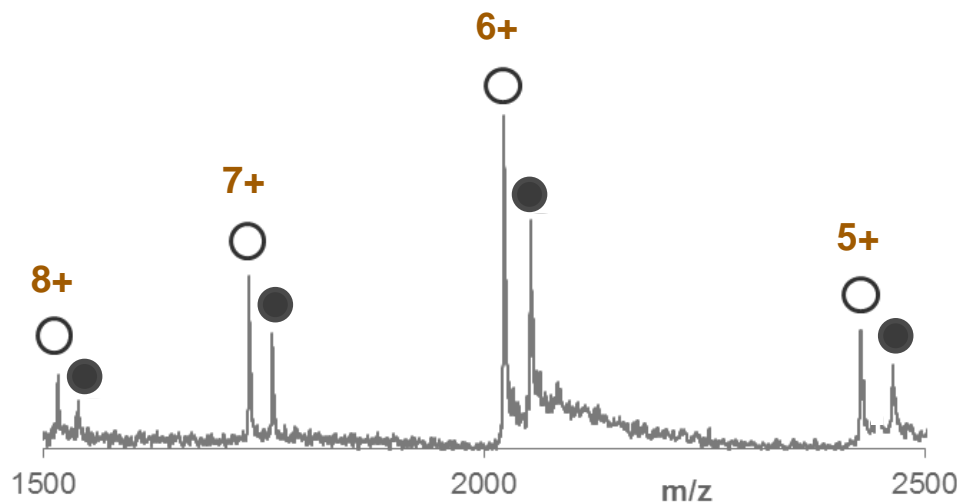
NCE = new chemical entity

## Orthogonal Screen for Hit Identification

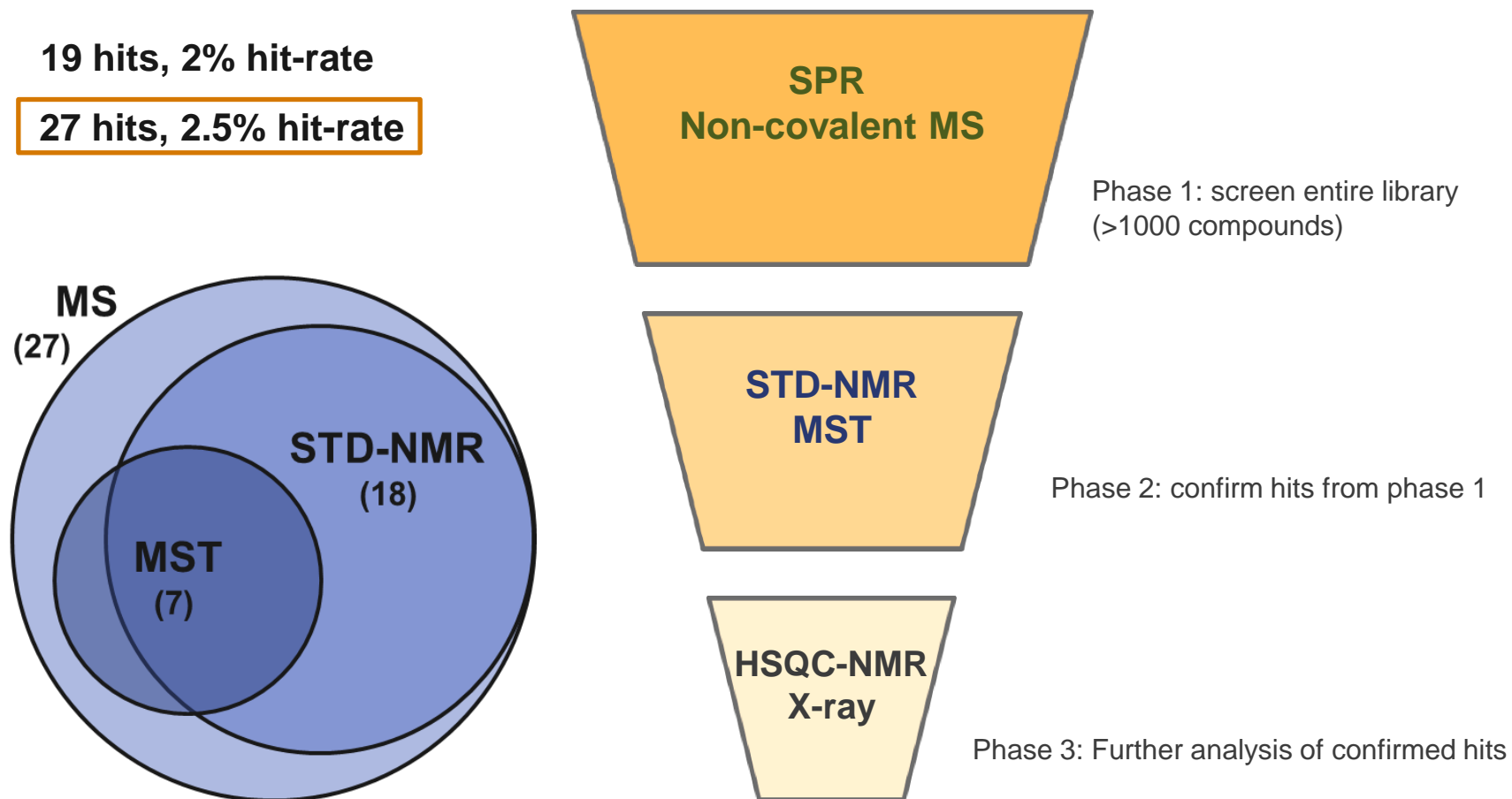
Library of 10s-1000s of fragments (~200 Da) or compounds (~500 Da)

Binding to purified protein target (10 kDa - 100 kDa): native-like state

Automated native MS protocol; ToF or Q-ToF MS



# Integrating Non-Covalent MS into In-House Screening Approach



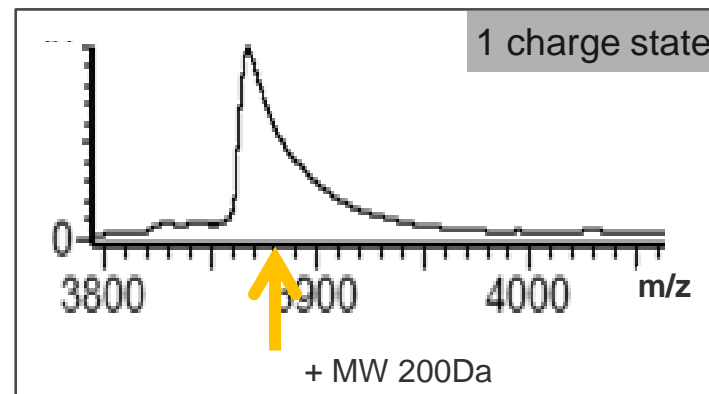
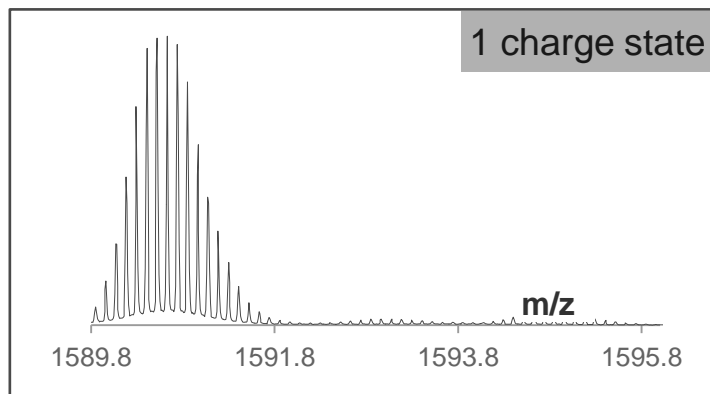
STD-NMR = Saturation Transfer Difference NMR  
MST = Microscale Thermophoresis

SPR = surface plasmon resonance

*Hannah Maple*

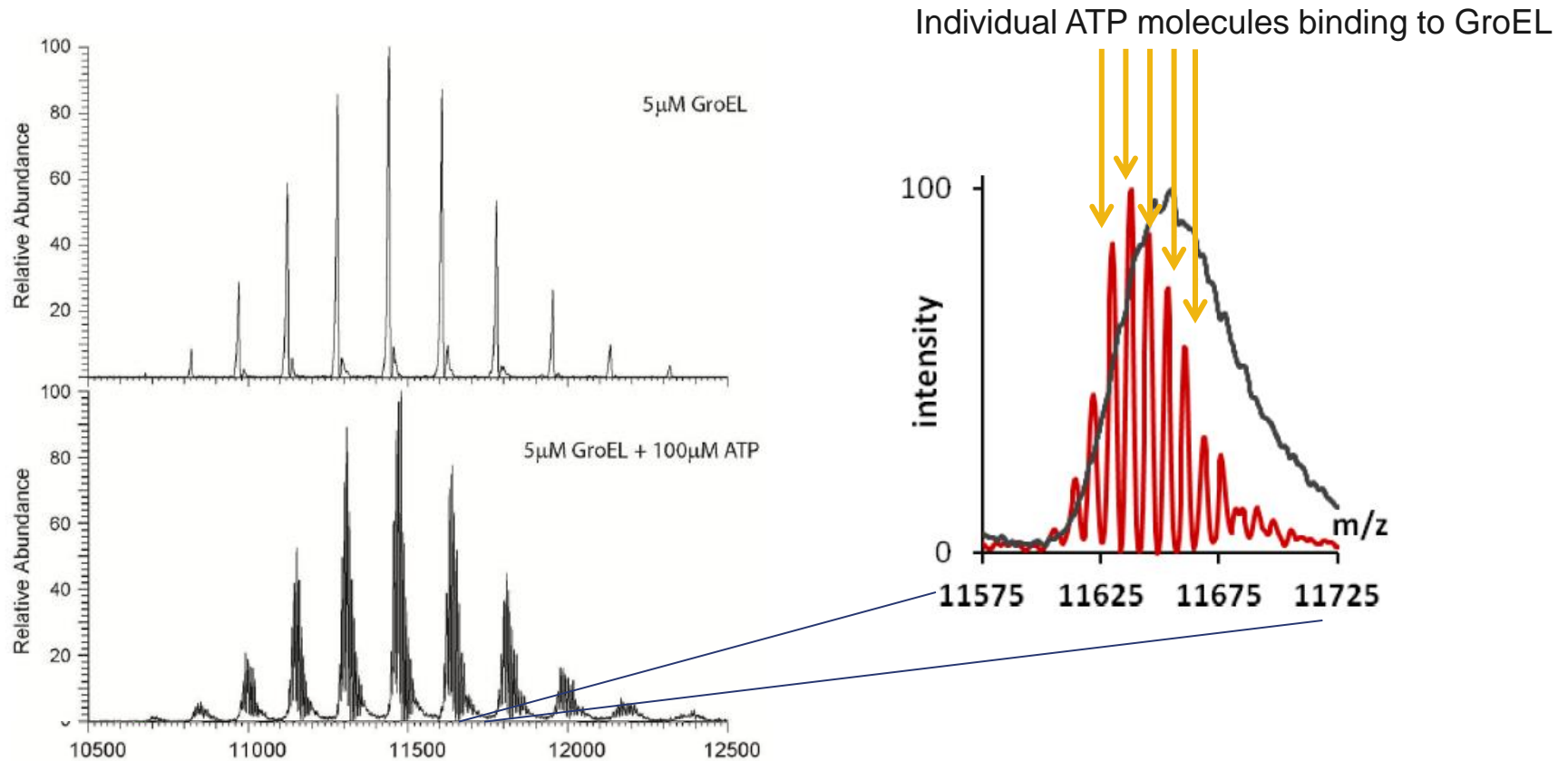
# Limitations of Screening by MS

- Measurement of binding limited by 'spectral resolution' - adducts, peak tailing, desalting  
[source conditions/ accelerating voltages must be kept to a minimum to retain compound binding]
- Speed/ Sensitivity:  
Medium throughput,  
Non-specific binding (ESI artefact, concentration)



# Orbitrap MS Analysis of Large Protein Complexes

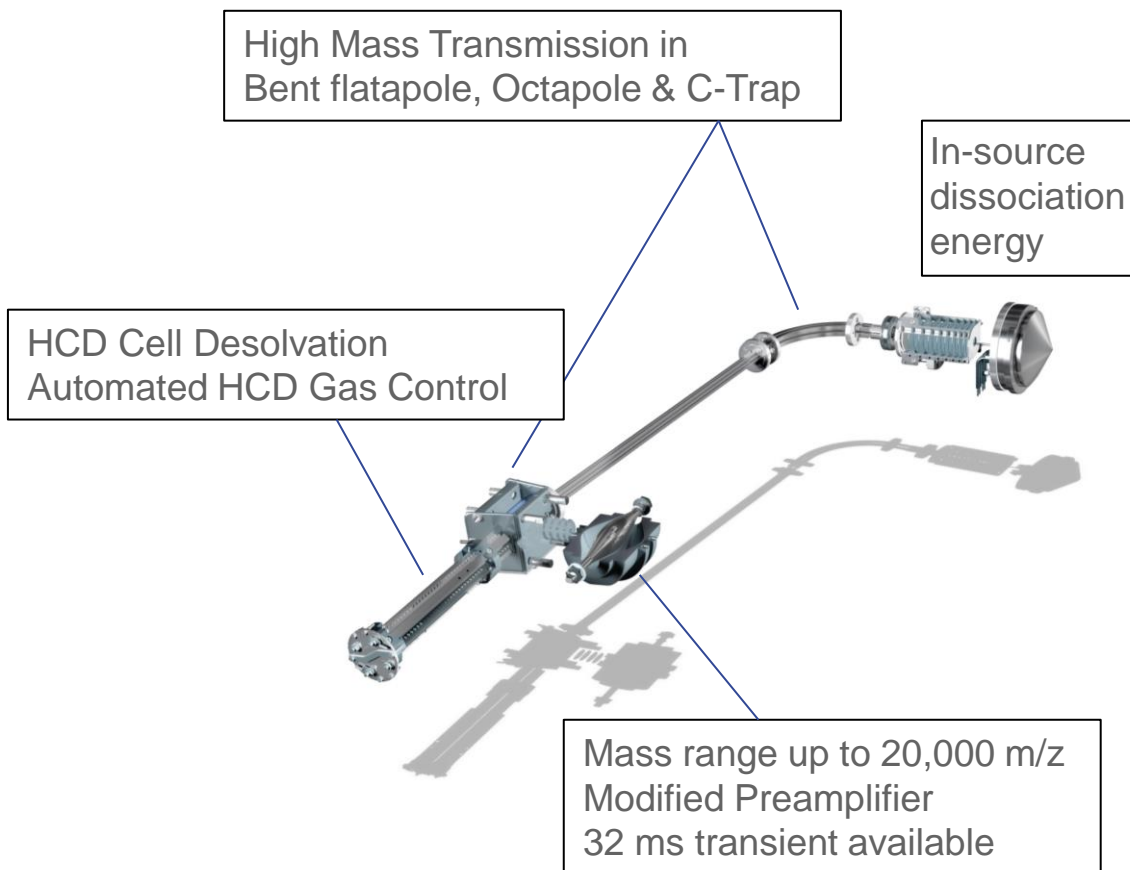
800 kDa GroEL + 507 Da ATP



TEST for protein-ligand analysis (smaller proteins, weak binding)  
COMPARE to existing technology

# Native Mass Spectrometry – Instrumentation

*Thermo Scientific™ Exactive™ Plus EMR Orbitrap mass spectrometer*





# Native Mass Spectrometry – Sample Introduction

*Thermo Scientific™ Exactive™ Plus EMR Orbitrap mass spectrometer*



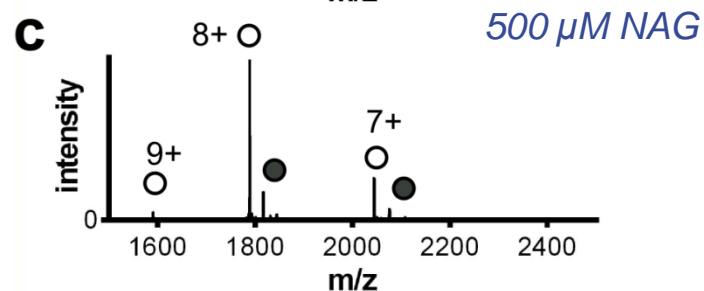
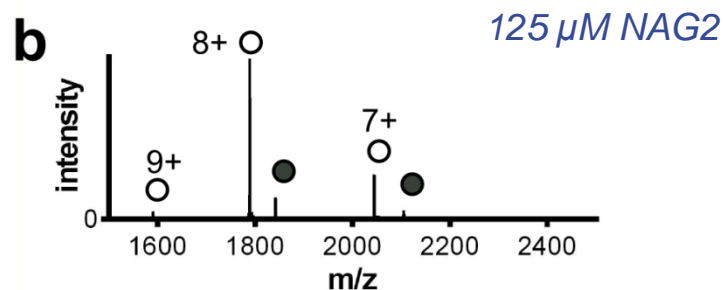
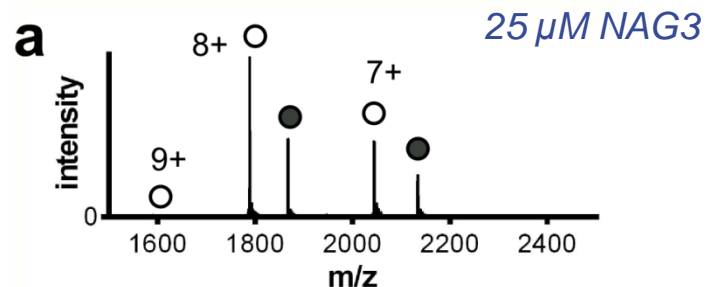
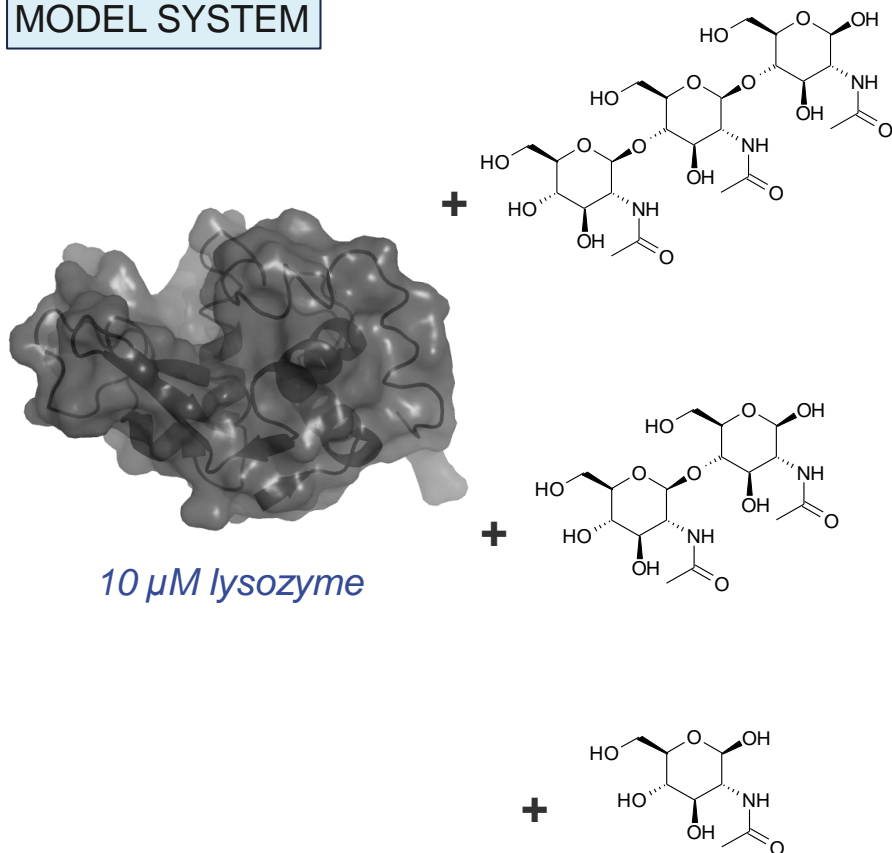
## Advion Triversa Nanomate

Automated chip-based nanoESI from 96- or 384- well plate



# Protein-Ligand Binding on Exactive Plus EMR MS

## MODEL SYSTEM



1 scan (< 1 sec)

10  $\mu$ M

100  $\mu$ M

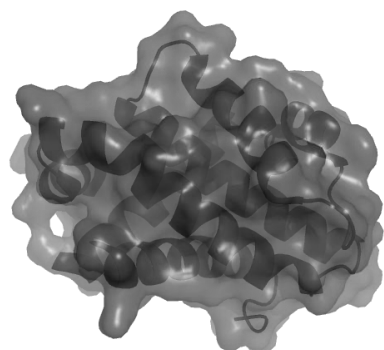
1 mM

$K_D$



# Protein-Ligand Binding on Exactive Plus EMR MS

DRUG DISCOVERY TARGET



10  $\mu$ M Bcl-xL

+

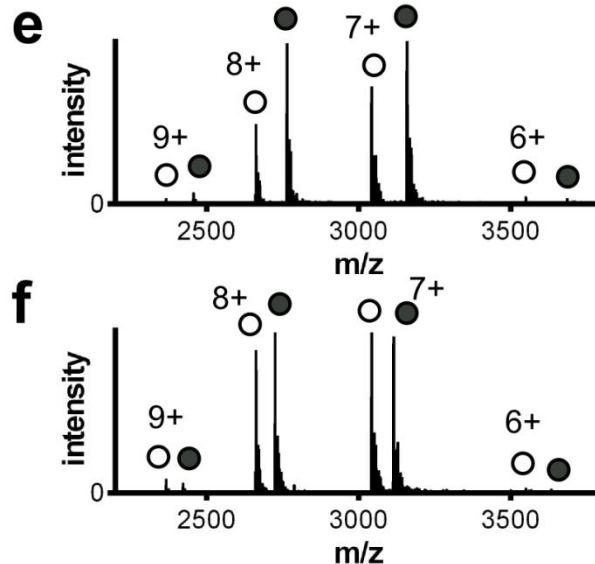
ABT737

Mw 812 Da

+

'cpd 1'

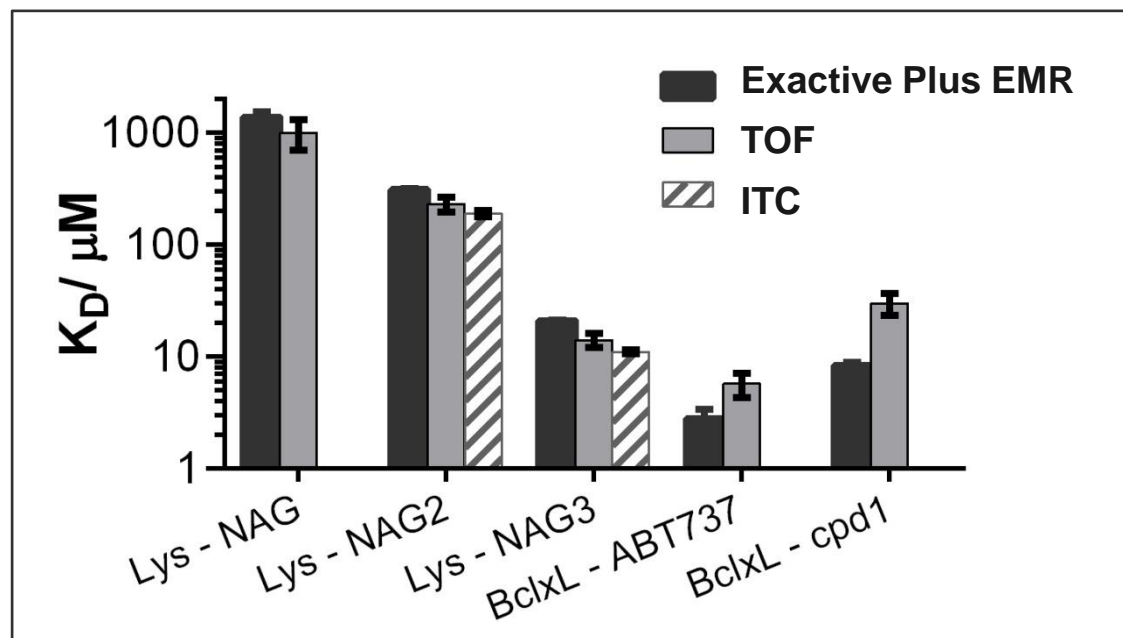
Mw 502 Da



1 scan (< 1 sec)

# Protein-Ligand Binding on Exactive Plus EMR MS

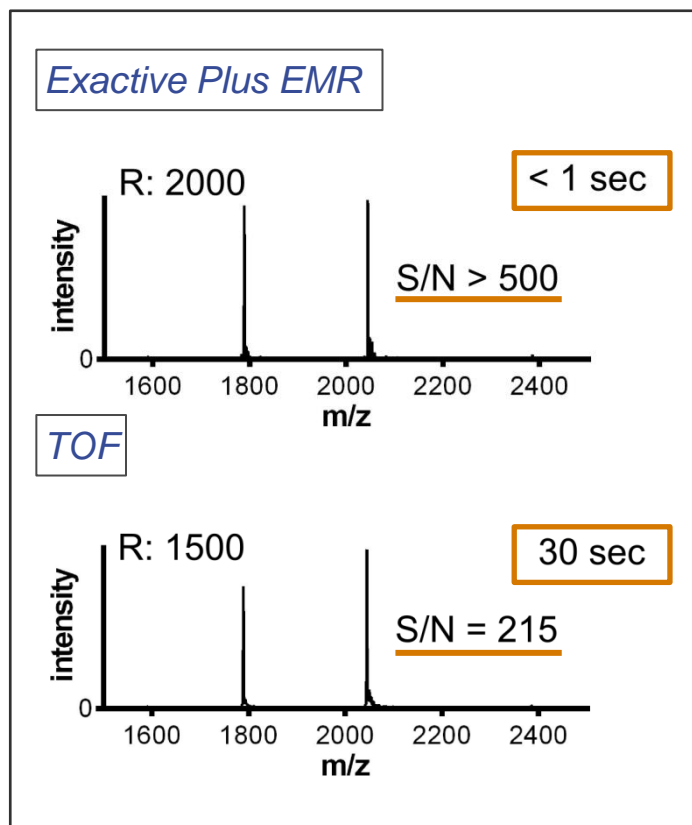
Estimation of binding strength, comparison with existing technology



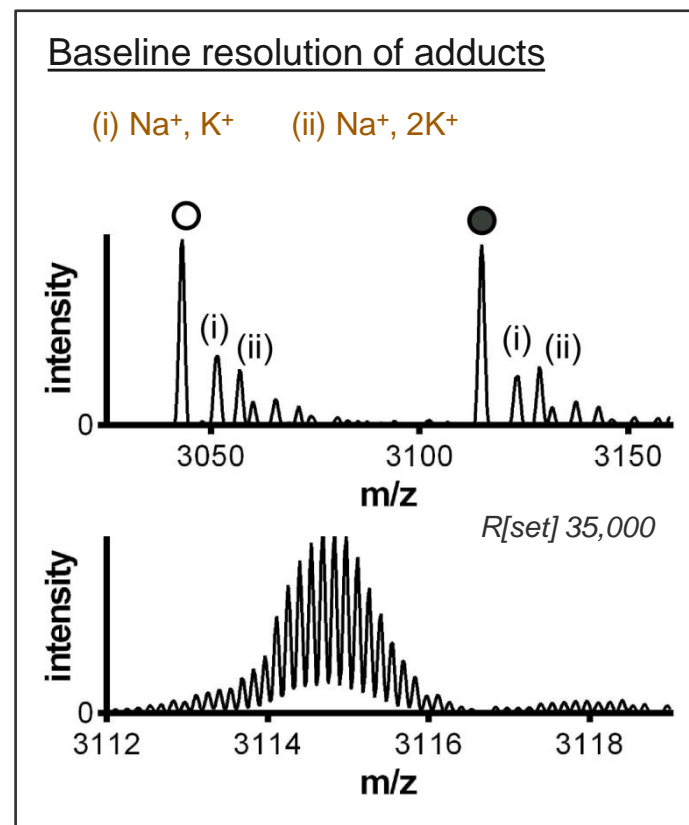
$K_D/\mu\text{M}$	E+ EMR	TOF	ITC
Lys-NAG	1375 ± 159	1000 ± 300	
Lys-NAG2	311 ± 7	230 ± 35	189 ± 11
Lys-NAG3	21 ± 0.4	14 ± 2	11 ± 0.4
BclxL- ABT737	2.8 ± 0.6	5.7 ± 1.4	
BclxL-cpd1	8.3 ± 0.5	30 ± 6.6	

# Sensitivity and Spectral Resolution

625 nM lysozyme

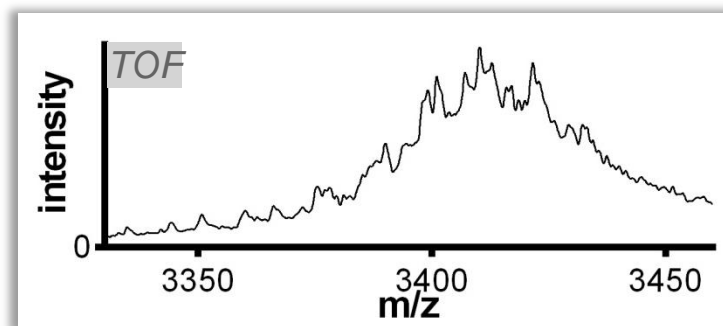
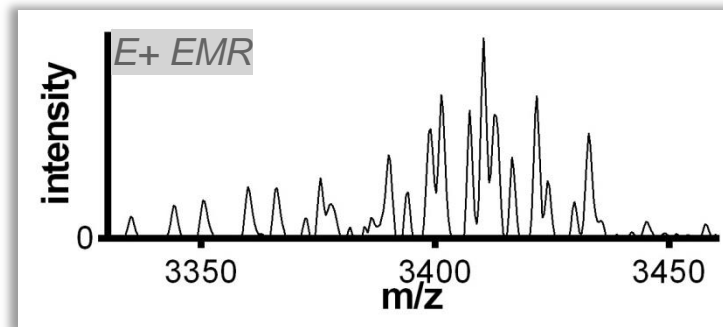
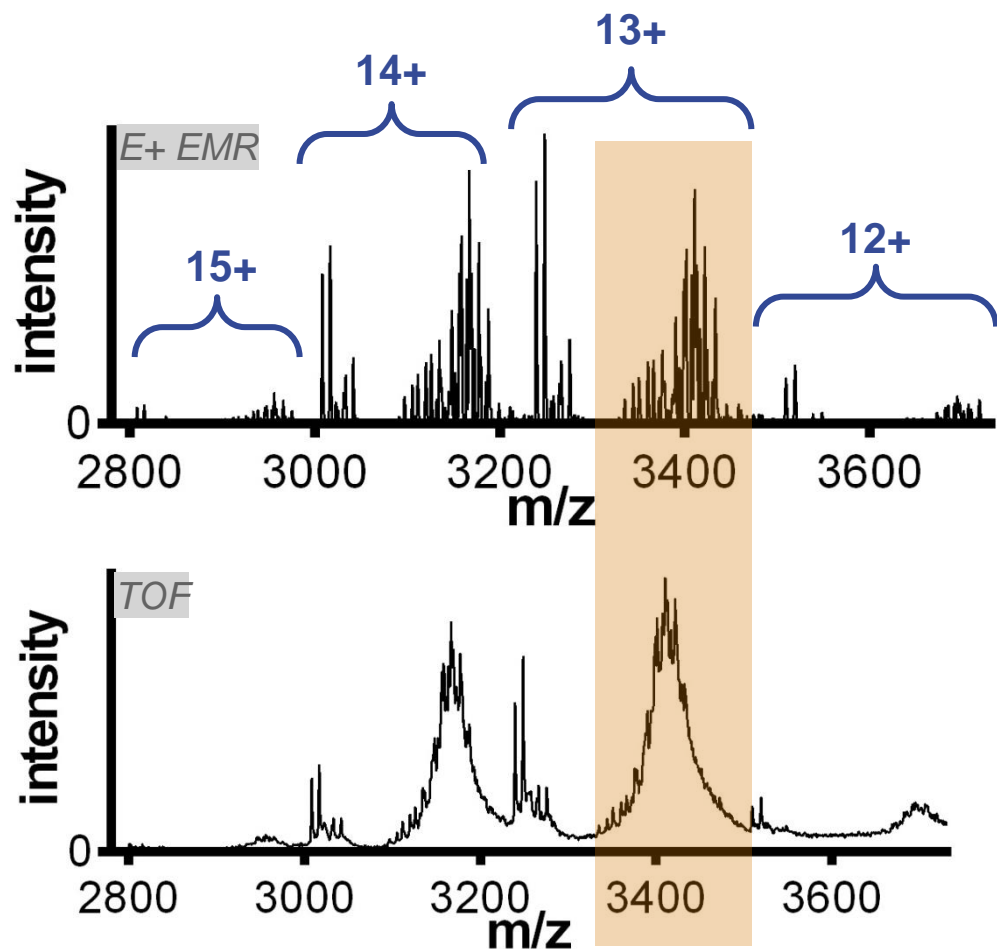


*BclxL*



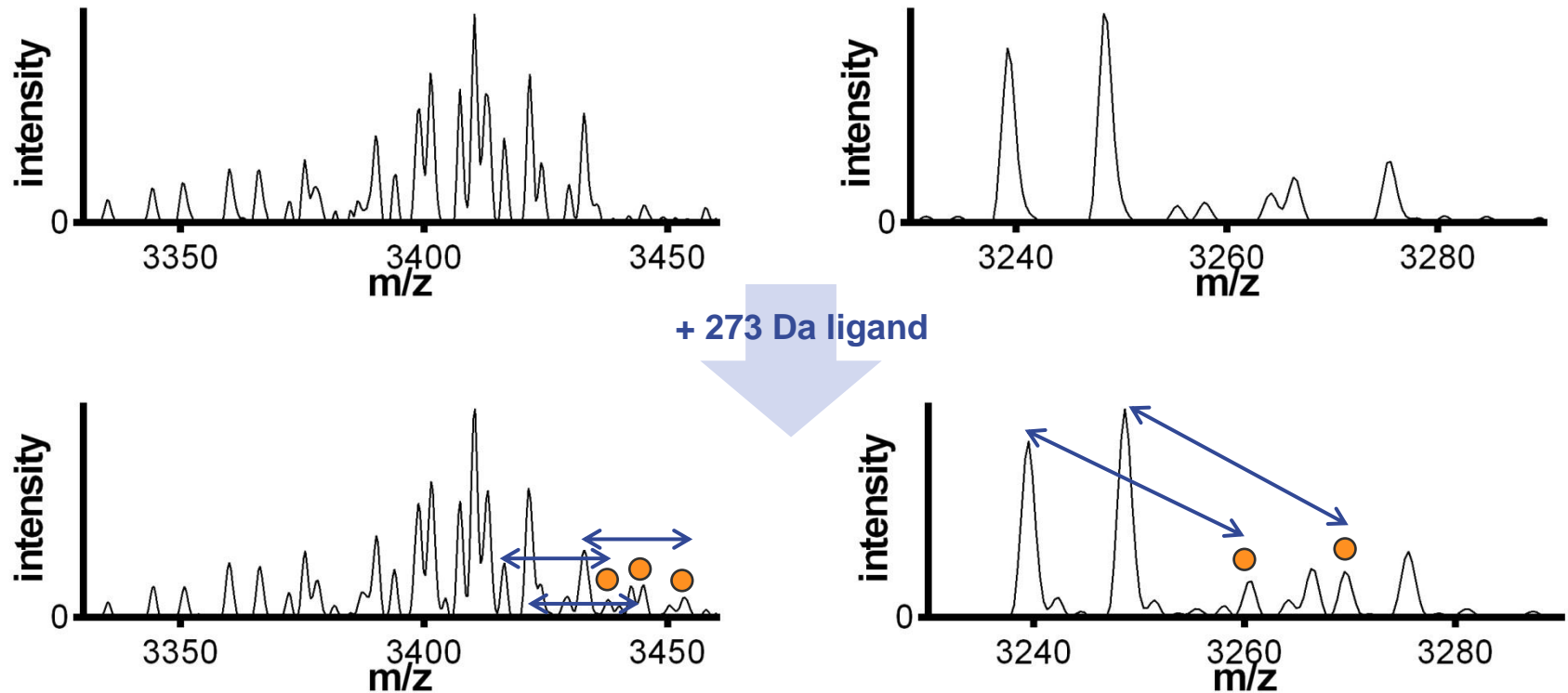
# Heterogeneous Proteins: Glycoproteins

40 kDa highly glycosylated protein:



# Measuring Fragments Binding to Glycosylated Proteins

Typical approach: enzymatic deglycosylation. May affect protein structure/ stability/ function.



Ligand binding observed directly with glycosylated protein

# Conclusions

**Non-Covalent MS can be used to screening small compounds and fragments for hit identification in drug discovery**

Evaluation of Exactive Plus EMR for this application (model systems):

Maintains non-covalent protein-ligand interactions during analysis ( $K_D$  to mM)

Spectra obtained within seconds - sensitive

Good spectral resolution, good peak shape

- ⇒ **More reliable measurement of binding of SMALL or WEAKLY-BINDING compounds**
- ⇒ **Faster analysis of interactions**
- ⇒ **Lower concentrations: reduced sample consumption, reduced non-specific artefacts**

# Acknowledgements

## UCB Celltech

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Phil Gilbert

### **MST**

Nicolas Basse

## Thermo Fisher Scientific

Olaf Scheibner  
Maciej Bromirski

## Advion Ltd

Mark Baumert  
Mark Allen

# Thank You!



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Driven by **science.**